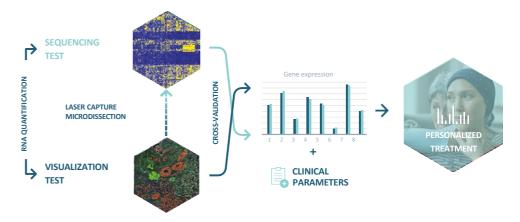
Multiplex8+ RESULTS



| PATIENT | SAMPLE | ORDERING PHYSICIAN |
|--------------|------------------------|--------------------|
| Name: | Specimen ID: MDX-PT-11 | Name: |
| ID: | Date of collection: | Address: |
| Report date: | Type: | Contact: |

TEST DESCRIPTION

The Multiplex8+ breast cancer test assesses RNA-based biomarkers by conducting a VISUALIZATION TEST that uses RNA fluorescent in situ hybridization (RNA-FISH) to visualize a panel of biomarkers. Based on the expression of these biomarkers and the tissue histology, laser capture microdissection is used to dissect out regions of interest. With these tumor-enriched samples, a SEQUENCING TEST that utilizes total RNA next generation sequencing to survey gene expression in a spatially resolved manner, is further carried out. Analytical validation of Multiplex8+ was conducted on a large retrospective cohort of 1 080 breast tumors.



THE TEST PROVIDES INFORMATION ABOUT:

- 1. RECEPTOR STATUS: for RNA expression of the estrogen receptor, progesterone receptor, Her2 receptor, and Ki67 measured and cross-validated by the two tests.
- **2. MOLECULAR SUBTYPE:** based on RNA gene expression tumor biology.
- **3. GENE SIGNATURES:** personalized for patients' tumor biology and clinical status.

A SUMMARY IS PROVIDED BELOW AND ADDITIONAL DETAILS ARE PROVIDED IN THE FOLLOWING PAGES.

RESULTS SUMMARY

RECEPTOR STATUS

| Sample | ESR1 | PGR | ERBB2 | MKI67 |
|--------|------|-----|-------|-------|
| Α | + | _ | + | _ |
| | | | | |

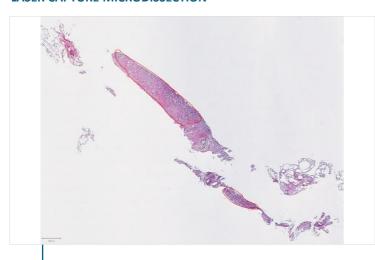
MOLECULAR SUBTYPE

| Intrinsic subtype | TNBC subtype |
|-------------------|--------------|
| Luminal A | - |
| | |

RELEVANT TREATMENT

| THERAPY | KEY FINDINGS | CLINICAL BENEFIT |
|-------------------|---|-------------------|
| Anti-Her2 | Gene expression, gene expression signature | Predicted benefit |
| Endocrine therapy | Gene expression, gene expression signature, molecular subtype | Predicted benefit |
| Chemotherapy | Chemotherapy Gene expression, gene expression signature | |

LASER CAPTURE MICRODISSECTION



Based on histological assessment and RNA-FISH biomarker expression, one sample (**Sample A**) was laser capture microdissected for further analysis.

RECEPTOR STATUS

| Sample | ESR1 | PGR | ERBB2 | MKI67 |
|--------|------|-----|-------|-------|
| Α | + | _ | + | _ |
| | | | | |
| | | | | |

Receptor status was determined using both the **VISUALIZATION TEST** and **SEQUENCING TEST**: the table shows results after cross-validation.

INTERPRETATION

 The results from both RNA-FISH and RNA-SEQ were consistent with the immunohistochemical findings.

MOLECULAR SUBTYPE

| Intrinsic subtype | TNBC subtype ²⁻⁴ |
|-------------------|-----------------------------|
| Luminal A | - |
| | |

Based on the **SEQUENCING TEST**, we used a consensus subtyping approach consisting of our proprietary 293 gene molecular subtyping signature, a research-based PAM50 test and the AIMS method to classify the intrinsic molecular subtype ¹. TNBC subtype, if applicable, was classified according to Lehmann ²⁻⁴.

INTERPRETATION

- The molecular classification of the subtype as Luminal A is inconsistent with the immunohistochemical and clinical designation. This is not uncommon. For example, in the METABRIC cohort, 18% of ER+/PR-/Her2+ samples were classified as Luminal A by PAM50.
- Luminal A tumors are characterized by expression of ER and/or PR and either negative or low expression of Her2 and the proliferation marker KI67. Luminal A tumors are low grade, have favorable prognosis, and respond well to endocrine therapies such as tamoxifen or aromatase inhibitors.

GENE SIGNATURE

• Based on the assigned molecular subtype, and TNBC subtype (if applicable), we evaluated several individual genes and gene signatures that demonstrate prognostic and predictive potential in early and advanced/metastatic settings.

| Treatment type/ Pathway | Gene signature | Description | Sample A Percentile | |
|----------------------------|--------------------------------------|---|---------------------|--|
| Prognosis | Consensus prognostic signature | The prognostic signature is derived from a consensus of three research-based prognostic signatures, including the 21-gene signature GENE21 5, the 70-gene GENE70 signature 6, and the 50-gene risk of relapse based on subtype alone (ROR-S) signature 7. The prognostic signatures are intended for early-stage breast cancer patients with ER+/Her2- IHC, lymph node-negative, or 1-3 positive lymph nodes. The score is reported as high, intermediate, or low. Patients with high signature scores are at a greater risk of relapse and may benefit from adjuvant chemotherapy, while patients with low scores have lower risk of relapse and may not benefit from adjuvant chemotherapy. | N/A | |

ID:

| Treatment | Gene | | Sample A |
|-----------------------|----------------------------|--|-----------------|
| type/ Pathway | signature | Description | Percentile |
| | ESR1 | The ESR1 and PGR genes encode for the estrogen (ER) and progesterone (PR) hormone receptors, respectively, which are involved in growth, metabolism, and | Medium (40%) |
| I | PGR | reproductive functions. High ER/PR is predictive of endocrine therapies and low or negative ER/PR is associated with poor prognosis ⁸ . | Low (21%) |
| Luminal signatures | ESR1_PGR average | The average gene expression of ESR1 and PGR. Higher levels of hormone receptors are predictive markers for endocrine therapies. | Medium (37%) |
| | E2F4_score | This gene signature assesses activity of the E2F4 transcription factor and its targets. A high E2F4 signature is associated with endocrine resistance to aromatase inhibitors and may predict sensitivity to CDK4/6 inhibitors ⁹ . | Low (12%) |
| | ERBB2 | The ERBB2 gene is translated into Her2, a receptor tyrosine kinase involved in cell growth/proliferation and is both a prognostic marker and predictive of response to Her2 targeted therapies ⁸ . | High (90%) |
| | MUC4 | Mucin 4 (MUC4) is a glycoprotein that is implicated in resistance to trastuzumab through interactions with the Her2 receptor. High MUC4 is associated with reduced sensitivity to trastuzumab 10 . | Low (25%) |
| Her2 | NRG1 | NRG1 codes for neuregulin 1, a ligand of the Her3 receptor. In the phase II NeoSphere trial, high NRG1 gene expression was associated with reduced response to neoadjuvant trastuzumab, but not combination trastuzumab-pertuzumab ¹¹ . | Medium (62%) |
| | pSTAT3-GS | A signature that predicts phosphorylation of STAT3 and was found to be predictive of trastuzumab resistance in the FinHer study ¹² . | Medium (47%) |
| | Her2 amplicon_ MDX | Proprietary MDX 43-gene signature used to assess Her2 status. | High (99%) |
| | Module7_ ERBB2 | Her2-signaling signature predictive of response to multiple anti-Her2 treatments in the I-SPY2 trial ¹³ . | High (100%) |
| | AURKA | Aurora Kinase A (AURKA) is a protein coding gene involved in cell proliferation and is an independent prognostic marker in breast cancer. | Low (8%) |
| Proliferation | MKI67 | MKI67 codes for the marker of proliferation Ki67 protein, a marker of poor prognosis in ER+/Her2– tumors, but not Her2+ or TNBC tumors. Ki67 levels are also predictive of sensitivity to neoadjuvant endocrine and chemotherapies ⁸ . | Low (1%) |
| . romeradon | Module11_ proliferation | Proliferation index used in I-SPY2 trial broadly predictive of pathological complete response in hormone receptor positive patients ⁴ . | Low (27%) |
| | Proliferation_ MDX | Proprietary MDX 7-gene signature used to assess cellular proliferation and cross-validate MKI67 expression levels. | Low (14%) |
| | CDK4 | Cyclin-dependent kinases 4 and 6 (CDK4 and CDK6) are important proteins that regulate cell cycle progression from G1 to S phases. They are the main targets of | High (75%) |
| | CDK6 | CDK4/6 inhibitors such as palbociclib (Ibrance), ribociclib (Kisqali), and abemaciclib (Verzenio); however, it is unclear whether their expression level predicts CDK4/6 inhibitor sensitivity. | Medium (60%) |
| CDK4/6 inhibitors | CCNE1 | | Low (26%) |
| | CCND3 | Elevated expression of the G1/S cell cycle regulators, CCNE1, CCND3, and CDKN2D, was associated with resistance to palbociclib (Ibrance) in the single-arm phase II neoadjuvant trial (NeoPalAna) 14. | Low (8%) |
| | CDKN2D | | Low (25%) |
| PIK3CA mutations | PIK3CA-GS | A gene signature that is predictive of mutations in the PIK3CA gene and consequently the PI3K inhibitor alpelisib (Piqray). A high PIK3CA-GS score is also associated with activation of the PI3K/AKT pathway and loss of mTORC1 signaling, which may be relevant for response to mTOR inhibitors (e.g., everolimus) ¹⁵ . | Low (28%) |

| Treatment type/ Pathway | Gene signature | Description | Sample A Percentile | |
|--|--|---|---------------------|--|
| | TOP1 | The gene encoding DNA topoisomerase I, an enzyme critical for DNA transcription, is a target for anticancer drugs. | High (88%) | |
| | TOP2A | The gene encoding DNA topoisomerase IIa, an enzyme critical for DNA transcription, is a target for anticancer drugs. | Low (16%) | |
| | RAD51 | The DNA repair protein RAD51 homolog 1 (RAD51) is involved in the repair of damaged DNA and is associated with resistance to chemotherapy. | Low (28%) | |
| | ERCC1 | The DNA excision repair protein ERCC-1 (ERCC1) is involved in the repair of DNA damage and is associated with resistance to chemotherapy. | High (69%) | |
| | TYMS | The Thymidylate Synthetase (TYMS) gene encodes a protein involved in DNA biosynthesis and is the target of the antimetabolite chemotherapy, 5-Fluorouracil ¹⁶ . | Low (2%) | |
| | SLC29A1 | SLC29A1 codes for the equilibrative nucleoside transporter 1 (ENT1) protein, which is a nucleoside transporter that is involved in transporting gemcitabine and capecitabine ¹⁷ . | Medium (40%) | |
| | DHFR | Dihydrofolate reductase is an enzyme coded by the DHFR gene and is involved in folate metabolism and cell growth. It is the target of the antimetabolite chemotherapy, methotrexate ¹⁸ . | Low (33%) | |
| | SLC19A1 | SLC19A1 codes for the reduced folate carrier 1 (RFC1) protein, which transports methotrexate into the cell ¹⁸ . | Low (27%) | |
| | CDK12 | The protein product of the Cyclin Dependent Kinase 12 (CDK12) gene regulates transcription, DNA repair pathways, and cell cycle ¹⁹ . | Medium (65%) | |
| Chemotherapy | MAPs_Mitotic_ki nases_neoadj_ch emo118 | A 118-gene signature predicting response to neoadjuvant taxane chemotherapy 20 . | Medium (35%) | |
| | MAPs_Mitotic_ki nases_neoadj_ch emo17 | A 17-gene signature predicting response to neoadjuvant taxane chemotherapy $^{20}.$ | Low (1%) | |
| | Early_Relapse_E R.Neg | Chemoresistance gene signature predicting early relapse in ER-negative (ER-) patients after taxane-anthracycline chemotherapy ²¹ . | Medium (62%) | |
| | Residual_ disease_ ER.Neg | Chemoresistance gene signature predicting residual disease in ER-negative (ER-) patients after taxane-anthracycline chemotherapy ²¹ . | High (68%) | |
| response_ER.Neg Early_RelaR.Pos Residual_disease_I | Pathologic_ response_ ER.Neg | Chemosensitivity gene signature predicting pathological complete response in ER-negative (ER-) patients after taxane-anthracycline chemotherapy ²¹ . | High (75%) | |
| | Early_Relapse_E R.Pos | Chemoresistance gene signature predicting early relapse in ER-positive (ER+) patients after taxane-anthracycline chemotherapy ²¹ . | Medium (50%) | |
| | Residual_ disease_ ER.Pos | Chemoresistance gene signature predicting residual disease in ER-positive (ER+) patients after taxane-anthracycline chemotherapy ²¹ . | Medium (50%) | |
| | Pathologic_ response_ ER.Pos | Chemosensitivity gene signature predicting pathological complete response in ER-positive (ER+) patients after taxane-anthracycline chemotherapy ²¹ . | Medium (65%) | |

ID:

| Treatment type/ | Gene signature | Description | Sample A | |
|-----------------------------------|---|---|-----------------|--|
| Pathway | 33.33 | | Percentile | |
| | PDCD1 | PDCD1 codes for the immune checkpoint marker PD-1. PD-1 is the target of pembrolizumab (Keytruda), an immunotherapy approved for the first-line treatment of metastatic TNBC. | Medium (50%) | |
| | CD274 | CD274 codes for the immune checkpoint marker PD-L1. PD-L1 is the target of atezolizumab (Tecentriq), an immunotherapy approved for approved for the first-line treatment of metastatic TNBC. | Medium (34%) | |
| | CTLA4 | Cytotoxic T lymphocyte-associated antigen 4 (CTLA4) is an immune checkpoint marker and the target of several immunotherapies such as durvalumab (Imfinzi). | Low (32%) | |
| Immune system | Module5_ TcellBcell | | Medium (47%) | |
| | Chemokine12 | Immune signatures predictive of response to pembrolizumab in TNBC | Low (26%) | |
| | STAT1 | patients enrolled in (I-SPY2 trial) ¹⁴ . All signatures, with the exception of Mast_cells, were associated with increased probability of achieving pathological complete response. | Medium (59%) | |
| | Dendritic_cells | | High (83%) | |
| | Mast_cells | | Low (11%) | |
| DNA damage and repair | VCpred_TN | DNA damage repair / immune signature predictive of response to veliparib (PARP inhibitor) and carboplatin (I-SPY2 trial) ¹⁴ . | Medium (63%) | |
| | VEGFA | A gene coding for vascular endothelial growth factor, a protein involved in angiogenesis, vasodilation, and endothelial cell growth. VEGF is the target of the drug bevacizumab (Avastin). | High (92%) | |
| Angiogenesis/ hypoxia | Hypoxia / Angiogenesis / Inflammatory_ MDX | Proprietary MDX 7-gene signature used to assess hypoxia, angiogenesis, and inflammation. Signature includes genes known to be predictive of response to bevacizumab (Avastin) in the neoadjuvant GeparQuinto trial ²² . | Medium (57%) | |
| | ERBB2 | ERBB2 codes for the protein receptor Her2, which is a target for classical anti- Her2 treatments. Low and ultralow levels of Her2 can be eligible for treatment with the antibody-drug conjugate, trastuzumab deruxtecan (Enhertu) ²³ . | High (90%) | |
| | TACSTD2 | TACSTD2 codes for Tumor-associated calcium signal transducer 2, also called Trop-2, which is the target of sacituzumab govitecan (Trodelvy), an antibody-drug conjugate approved for metastatic TNBC 24 . | Medium (60%) | |
| | NECTIN4 | Nectin Cell Adhesion Molecule 4 (NECTIN4) is a cell adhesion molecule that is a target for antibody-drug conjugates in clinical trials for breast cancer. | High (78%) | |
| ADC (antibody- drug conjugate) | ERBB3 | ERBB3 codes for a member of the epidermal growth factor receptor (EGFR) family of receptor tyrosine kinases. It is under investigation in clinical trials for the antibody-drug conjugate patritumab deruxtecan. | Low (32%) | |
| targets | FOLR1 | FOLR1 encodes the protein Folate Receptor Alpha, which is an antibody-drug conjugate target under investigation for the treatment of metastatic TNBC in several phase 1 and 2 clinical trials. | Low (16%) | |
| | F3 | F3 codes for tissue factor, coagulation factor III a target of several antibody- drug conjugates in phase 1 clinical trials. | Medium (62%) | |
| | SLC39A6 | The SLC39A6 genes encodes for the zinc transporter LIV-1, which is highly expressed in luminal breast cancers and is under investigation in several phase 1 and 2 clinical trials. | Low (0%) | |
| | TPBG | The trophoblast glycoprotein (TPBG) is overexpressed in many breast cancers and is the target of at least two antibody-drug conjugates undergoing phase 1 clinical trials. | Medium (57%) | |

| Treatment type/ Pathway | Gene signature | Description | Sample A Percentile | |
|----------------------------|----------------|---|------------------------|--|
| | ROR2 | A gene that encodes the Receptor Tyrosine Kinase Like Orphan Receptor 2 protein, a target of the antibody-drug conjugate (Ozuriftamab Vedotin (BA3021/CAB-ROR2-ADC) that is under investigation in a phase clinical trial for advanced solid cancers, including TNBC. | Low (32%) | |
| | CD276 | This gene codes for an immune checkpoint marker called CD276 (also known as B7-H3). It is the target of the antibody-drug conjugate (Mirzotamab clezutoclax (ABBV-155) that is in a phase 1 and 2 clinical trial for advanced solid cancers, including breast cancer. | High (69%) | |
| | VTCN1 | V-Set Domain Containing T Cell Activation Inhibitor 1 (VTCN1 also called B7-H4) is an immune checkpoint marker and the target of the antibody-drug conjugate, SGN-B7H4V, which is under investigation in a phase1 clinical trial for advanced solid cancers, including breast cancer. | Medium (60%) | |
| | CEACAM5 | A gene that encodes CEA Cell Adhesion Molecule 5 protein, a target of the antibody-drug conjugate Tusamitamab ravtansine (SAR408701) that is under investigation in a phase 2 clinical trial for advanced solid cancers, including breast cancer. | High (95%) | |

INTERPRETATION AND RECOMMENDATIONS

- The elevated levels of ERBB2 and related Her2 amplicon gene signatures together with low-moderate expression of resistance markers (MUC4, NRG1 and pSTAT3-GS) suggest Her2-related treatments like trastuzumab may be beneficial.
- High ERBB2 and TOP1, which is the target of the cytotoxic payload for the ADC trastuzumab deruxtecan suggest this may be beneficial.
- The classification of a luminal A subtype, moderate ESR1 expression, and low E2F4 score suggests endocrine therapies like tamoxifen and aromatase inhibitors may be beneficial.
- The tumor has several markers of resistance to chemotherapies such as 5-fluorouracil (low TYMS expression), gemcitabine/capecitabine (moderate SLC29A1 expression), methotrexate (low SLC19A1 expression), low proliferation, and low (MAPs_Mitotic_kinases_neoadj_chemo17) and moderate (Pathologic_response_ER.Pos) expression of signatures predictive of anthracycline/taxane-based chemotherapies, suggesting uncertain benefits of chemotherapy.
- This sample shows high expression of antibody-drug conjugate target CEACAM5 (95th percentile), which is under investigation in phase 2 of clinical trial.

REFERENCES

1. Gendoo, D.M.A. et al. Bioinformatics 32(7): 1097–1099 (2016). 2. Lehmann, B. D. et al. J Clin Invest 121: 2750–2767 (2011). 3. Lehmann, B. D. et al. PLoS One 11: e0157368 (2016). 4. Bareche, Y. et al. Ann Oncol 29: 895–902 (2018). 5. Paik, S. et al. N Engl J Med 351(27): 2817-2826 (2004). 6. van't Veer, L.J. et al. Nature 415(6871): 530-536 (2002). 7. Parker, J.S. et al. J Clin Oncol 27(8): 1160-1167 (2009). 8. Cardoso, F. et al. Ann Oncol 30(8): 1194-1220 (2019). 9. Guerrero-Zotano, A.L. et al. Clin Cancer Res 24(11): 2517-2529 (2018). 10. Mercogliano, M.F. et al. Clin Cancer Res 23(3): 636-648 (2017). 11. Guardia, C. et al., Clin Cancer Res 27(18): 5096-5108 (2021). 12. Sonnenblick, A. et al. BMC Med 13:177 (2015). 13. Wolf, D. M. et al. Cancer Cell 40: 609-623.e6 (2022). 14. Ma, C.X. et al. Clin Cancer Res 23(15): 4055-4065 (2017). 15. Loi, S. et al. PNAS 107(22): 10208-10213 (2010). 16. Foekens, J.A. et al. Cancer Res. 61: 1421-1425 (2001). 17. Mackey, J.R. et al. Clin Cancer Res. 8(1): 110-116 (2002). 18. Yang, V. et al. RSC Med Chem. 11(6): 646-664 (2020). 19. Filippone, M.G. et al. Nat Commun. 13(1): 2642 (2022). 20. Rodrigues-Ferreira, S. et al. Proc Natl Acad Sci USA 116(47): 23691-23697 (2019). 21. Hatzis, C. et al. JAMA 305(18):1873-81 (2011). 22. Karn, T. et al. Clin Cancer Res 26: 1896–1904 (2020). 23. Modi, S. et al. N Engl J Med 387: 9–20 (2022). 24. Michaleas, S. et al. ESMO Open 7 (2022).